



## Synthesis and Influenza Virus Sialidase Inhibitory Activity of the 5-Desacetamido Analogue of 2,3-Didehydro-2,4-dideoxy-4-guanidiny-N-acetylneuraminic acid (GG167)

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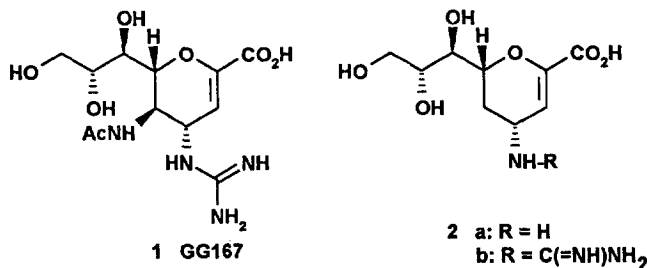
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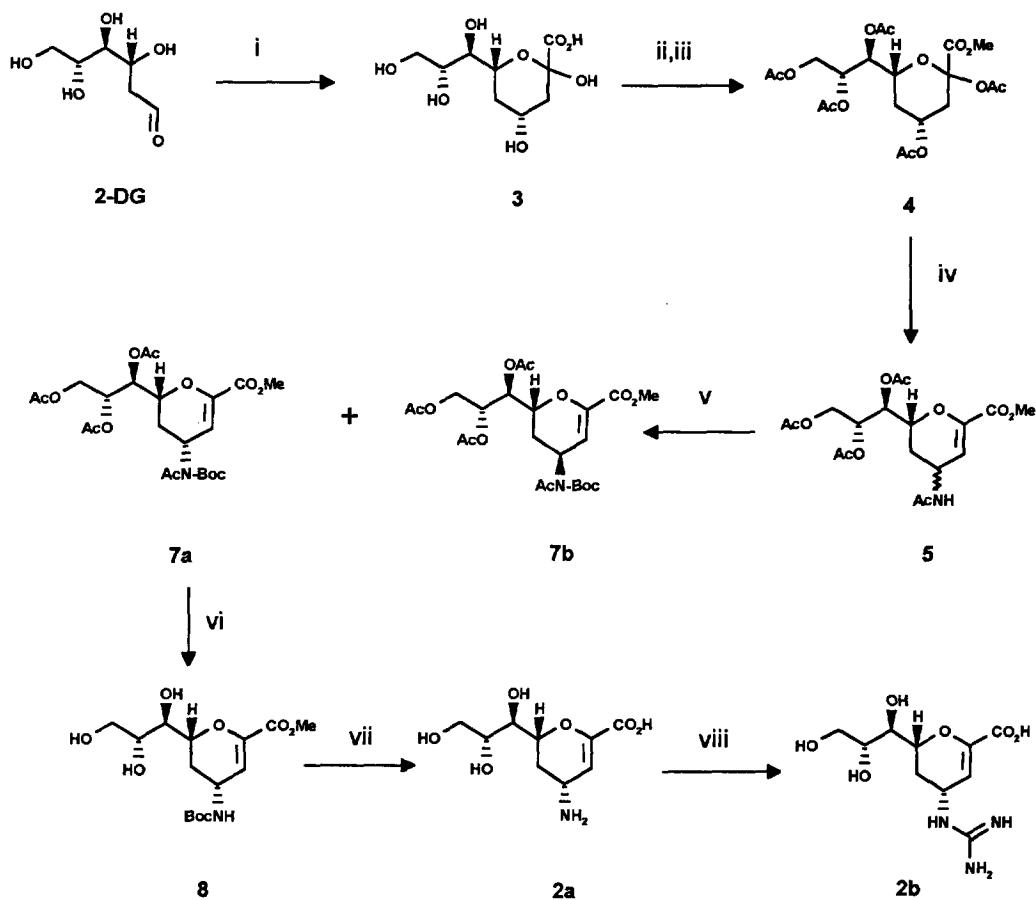
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**Abstract:** The title compound **2b** has been synthesised in 8 steps from 2-deoxy-D-glucose (2-DG). Key transformations were an aldolase reaction utilising 2-DG to form the 9 carbon sugar **3**, and then introduction of the required unsaturation and a 4-amino substituent in a one-pot process. The poor inhibitory activity of **2b** demonstrates that the 5-acetamido substituent in 2,3-didehydro-2,4-dideoxy-4-guanidiny-N-acetylneuraminic acid (GG167) **1** is critical for sialidase binding.

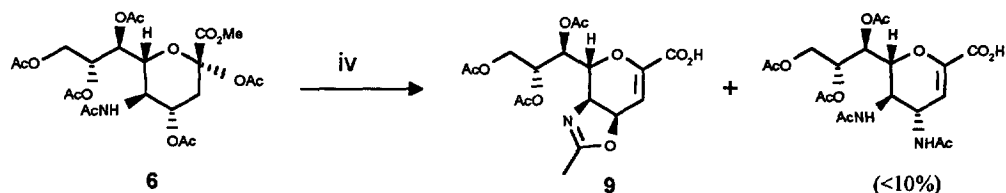
2,3-Didehydro-2,4-dideoxy-4-guanidiny-N-acetylneuraminic acid (GG167) **1** is a potent and selective inhibitor of influenza virus sialidase. It was rationally designed from the X-ray crystal structure of sialic acid bound to influenza sialidase with the aid of molecular modelling and computational chemistry techniques.<sup>1,2</sup> Recently the synthesis of **1** from N-acetyl-D-neuraminic acid (NANA) has been reported.<sup>3-6</sup> Formally, this represents a synthesis of **1** from N-acetyl-D-mannosamine (NAM) since NANA is available from NAM and pyruvate using an aldolase reaction.<sup>7</sup> In order to understand the contribution to sialidase binding made by each of the groups on the dihydropyran ring, we and others have embarked on the synthesis of analogues of **1**.<sup>6,8,9</sup> In this communication, we describe the synthesis and biological properties of **2a** and **2b** which lack a substituent at the 5-position of the dihydropyran ring. This has been achieved by utilising 2-DG as substrate in the aldolase reaction,<sup>7</sup> and subsequently introducing the required unsaturation and a 4-amino substituent in a one pot process involving an unexpected Ritter reaction.<sup>10,\*</sup>





**Reagents and conditions:** i) Aldolase, sodium pyruvate (41%) ii) MeOH / Dowex ( $H^+$ ) (90%) iii)  $Ac_2O$ , Pyridine, DMAP (75%) iv) TMS-OTf, MeCN (2:1  $\beta$ : $\alpha$ , 80%) v)  $Boc_2O$ , DMAP, dioxan, separate isomers vi) NaOMe, MeOH vii) NaOH aq then TFA/ $CH_2Cl_2$  (95% over 3 steps) viii) bis-Boc-PCH then TFA/ $CH_2Cl_2$ .

Scheme 1



Scheme 2

Immobilised NANA-aldolase<sup>11</sup> catalysed the conversion of 2-DG to the key intermediate **3** (Scheme 1). Typically, bioconversions were carried out at 20°C in a 2L flask (250 ml working volume) containing 2-DG (25 g) and sodium pyruvate (69 g). The pH was adjusted to 7.5 with NaOH and reactions were started by the addition of washed immobilised beads (150 g wet weight). Upon completion of the reaction, enzyme beads were removed by filtration and washed with distilled water (250 ml). The combined filtrate (500 ml) was passed through a bed (400 ml) of Amberlite 200 (H<sup>+</sup>) to remove sodium ions. The eluate was processed through a bed (500 ml) of Macro-Prep High Q on which 2-DG was not retained. The C-9 sugar **3** was then eluted with 0.5M acetic acid. Finally, the solution containing product was evaporated to low volume and then to dryness in the presence of 2-propanol to afford **3**.

Following established procedures, the carboxyl group in **3** was esterified using methanol and the hydroxyl groups peracetylated to give the fully protected intermediate **4**. Using methodology developed for the synthesis of **1**, compound **4** was next treated with trimethylsilyl triflate (TMSOTf) in acetonitrile.<sup>6</sup> In this reaction, whilst expecting elimination of the anomeric acetate, we also observed clean substitution of the newly formed allylic acetate by MeCN *via* a Ritter reaction to give a mixture of 4-acetamides **5** (2:1 ratio). A similar Ritter reaction was observed as a minor side reaction when compound **6** was subjected to the same conditions, although in this case the major product of the reaction was the oxazoline **9** formed *via* intramolecular attack at C-4 by the 5-acetamido group (Scheme 2).<sup>6,12</sup>

The individual isomers of **5** could not be separated at this stage, but after N-Boc protection, the bis-N-acylated derivatives **7a** and **b** were readily separated and the pure  $\alpha$ -isomer **7a** (minor product) was further elaborated to the targets **2a** and **2b**. Thus, the major isomer produced in the Ritter reaction resulted from inversion of the stereochemistry of compound **4** at the C-4 position.<sup>13</sup>

The final stages of the synthesis were completed using standard reactions. Thus, deacetylation of **7a** with methanolic sodium methoxide afforded the triol **8**. This was deprotected by treatment with aqueous sodium hydroxide followed by TFA/CH<sub>2</sub>Cl<sub>2</sub> to afford the amine **2a**. Guanylation of **2a** proceeded smoothly using N,N'-bis-Boc-1H-pyrazole-1-carboxamidine (bis-Boc-PCH)<sup>14</sup> and subsequent TFA deprotection afforded pure desacetamido GG167 **2b**.<sup>15</sup>

Compounds **2a** and **b** were evaluated as inhibitors of influenza A and B sialidase.<sup>16</sup> Both compounds displayed markedly reduced affinity for the viral enzymes when compared to **1** (Table 1).

**Table 1: Inhibition of Influenza A and B sialidases by compounds 1, 2a & 2b<sup>17</sup>**

<u>Compound</u>	<u>IC<sub>50</sub> (μM)</u>	
	<u>Flu A (Aichi)</u>	<u>Flu B (Victoria)</u>
<b>1</b>	0.005	0.004
<b>2a</b>	>500	-
<b>2b</b>	130	>400

In conclusion therefore, using a combination of enzyme mediated aldol chemistry and a novel one pot reaction for elimination and introduction of a nitrogen substituent, we have prepared **2a** and **2b** which are analogues of 2,3-didehydro-2,4-dideoxy-4-guanidiny-N-acetylneuraminic acid (GG167) lacking a 5-substituent on the dihydropyran ring. Subsequent evaluation of these compounds against influenza virus

sialidase has established the critical importance of the 5-acetamido group for good binding affinity of GG167.

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15. **Compound 2a:**  $^1\text{H}$  NMR ( $\text{D}_2\text{O}$ ): 6.04 (1H, dd, H-3, J 5.1, 1.7 Hz), 4.40 (1H, ddd, H-6, J 12.4, 2.3, 1.9 Hz), 4.15 (1H, m, H-4, J 5.8, 4.8, 1.5 Hz), 3.90 (1H, m, H-8, J 9.0, 6.1, 2.9 Hz), 3.82 (1H, dd, H-9, J 12.0, 2.9 Hz), 3.63 (1H, dd, H-9', J 12.0, 6.1 Hz), 3.56 (1H, dd, H-7, J 9.0, 1.9 Hz), 2.31 (1H, m, H-5 pseudo axial J 15.4, 12.4, 5.8 Hz), 2.03 (1H, dm, H-5 pseudo equatorial J 15.4, 2.3, 1.7, 1.5 Hz). High Res MS: Found  $\text{MH}^+$  234.09772, calcd for  $\text{C}_9\text{H}_{16}\text{NO}_6$  234.09776.  
**Compound 2b:**  $^1\text{H}$  NMR ( $\text{D}_2\text{O}$ ): 5.97 (1H, bd, H-3), 4.58 (1H, ddd, H-6, J 13, 7.2, 2 Hz), 4.44 (1H, m H-4), 3.93 (1H, ddd, H-8, J 9, 7, 2.6 Hz), 3.88 (1H, dd, H-9, J 14, 2.6 Hz), 3.63 (2H, m, H-7,9), 2.23 (1H, dm, H-5 pseudoequatorial, J 13, 6.5, 1.8 Hz), 1.92 (1H, br t, H-5, pseudo axial, J 13 Hz).  $\nu_{\text{max}}$  (KBr) 3600-3200 (br), 1668, 1612, 1202, 1145  $\text{cm}^{-1}$ . High Res MS: Found  $\text{MH}^+$  276.11928, calcd for  $\text{C}_{10}\text{H}_{18}\text{N}_3\text{O}_6$  276.11956.
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\*. *Note added in proof:* Since the submission of this manuscript a report has appeared which describes a Ritter reaction on a heptulose sugar derivative similar to the one which we observe in the conversion  $4 \rightarrow 5$ . Driguez P-A, Barrere B, Quash G and Doutheau A. *Carbohydr Res.* **1994**, *262*, 297.